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# *Mind the gap between non-activated (non-aggressive) and activated (aggressive) indoor fungal testing: impact of pre-sampling environmental settings on indoor air readings*

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## **Abstract**

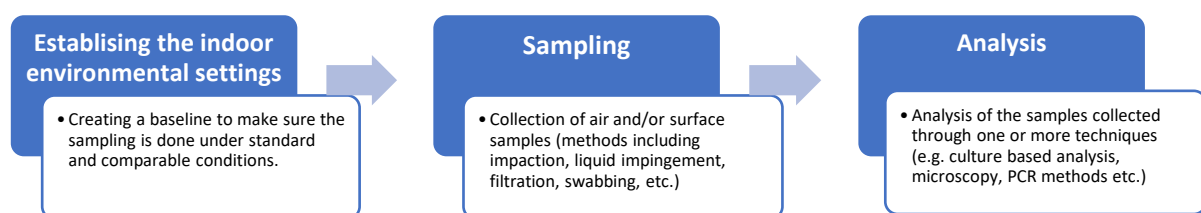
Indoor fungal testing has been within the researchers' scope of interest for more than a century. Various sampling and analysis techniques have been developed over the years, but no testing protocol has been yet standardised and widely accepted by the research and practitioner communities. The enormous diversity in fungal taxa within buildings with varied biological properties, and implications on the health and wellbeing of the occupants and the building fabric complicates the decision-making process for selecting an appropriate testing protocol. This study aims to present a critical review of non-activated and activated approaches to indoor testing, with an emphasis on the preparation of the indoor environment prior to sampling. The study demonstrates the differences in the outcomes of non-activated and activated testing through a set of experimental work in idealised conditions and a case study. The findings suggest that larger particles are particularly sensitive to the sampling height and activation, and that non-activated protocols, despite dominating the current literature, can significantly underestimate the fungal biomass and species richness. Therefore, this paper calls for better-defined and activated protocols that can enhance robustness and reproducibility across the research domain focused on indoor fungal testing.

**Keywords:** Mould; Fungal testing; Particles; Indoor fungi; Activated; Non-activated; Mould growth assessment

## **1.0 Introduction**

Indoor fungal growth may affect the health of occupants (WHO, 2009; 2013), disturb their comfort and well-being, and lead to damage to the building fabric (Anderson, 1998; Singh, 1999). Therefore, it is of critical importance to be able to measure the extent of fungal growth correctly in a given indoor environment. Through testing, researchers aim to quantify fungal biomass, determine the conditions under which fungi flourish, and assess whether the property needs remediation (Heinsohn 2007). However, though many sampling and analysis techniques have been proposed and widely implemented (BS EN ISO 16000-19:2014), indoor fungal testing has not yet been fully standardised through well-established protocols (Aktas, 2018a; Aktas et al, 2020).


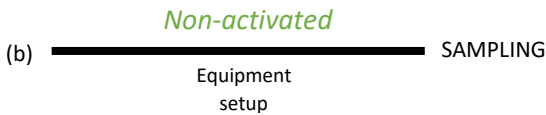
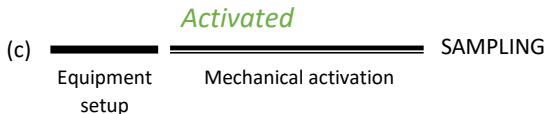
Indoor fungal testing protocols are typically composed of the following steps: (1) establishing environmental settings prior to sampling, (2) collection of samples and (3) sample analysis via one or more techniques to estimate the amount and/or the contents of indoor fungal flora (Figure 1).



**Figure 1: Schematic representation of the main stages of indoor fungal testing**

The first stage of testing includes all the activities carried out prior to sampling i.e., the establishment of the indoor environmental settings. The preparation may be done by creating a completely still environment by restricting any access to or movements in the spaces to be tested for some time prior to testing (termed as non-activated sampling) or by resuspending particles through mechanical means such as an air blower for a predetermined amount of time, often defined as a function of the room surface area or volume (termed as activated sampling). Between these two extremes are the protocols where the equipment setup and some movement of the occupants or the investigators prior to the testing are allowed (Table 1).

**Table 1: Some examples of non-activated (a & b) and activated protocols (c) [length of line segments not indicative of the durations associated to their labelled activities relative to each other]**

Examples of different protocols	Example studies
<p>(a) <b>Non-activated</b></p> 	<p>Terčelj et al, 2011: Windows and doors were closed for several hours prior to sampling. Shinohara et al, 2021: Tests were carried out in unoccupied spaces.</p>
<p>(b) <b>Non-activated</b></p> 	<p>Gent et al, 2012, Dallongeville et al, 2015; Cahna et al. 2015; Caballero et al, 2021: Activities prior to sampling were not restricted. The rooms were occupied before and during the sampling but no mechanical activation was carried out</p>
<p>(c) <b>Activated</b></p> 	<p>Aktas et al 2018 (a-c): Mechanical activation for approx. 1min/10m<sup>2</sup> was carried out after the equipment setup. Upon completion of the activation, sampling was initiated.</p>

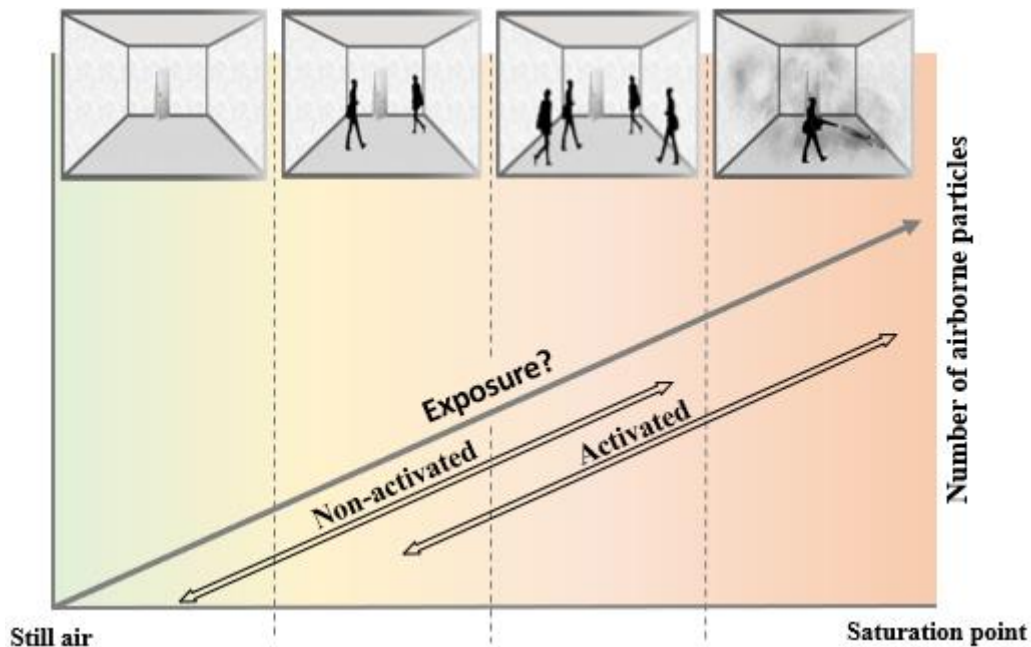
As seen, while there is substantial literature on the selection of sampling and analysis techniques depending on the aim of the investigation and the availability of tools (Heinsohn, 2007), the standardization of protocols with regards to the preparation of the indoor environment prior to sampling has gained limited attention and interest (Buttner and Stetzenbach, 1993; Rao, Burge & Chang, 1996; Flannigan, 2011). This situation manifests as additional complexity when comparing and contrasting findings of multiple research studies, even when the same sampling and analysis techniques are used.

### 1.1. Non-activated (non-aggressive) and activated (aggressive) protocols

The preparation of the environmental settings prior to sampling is often described by different and conflicting terms used interchangeably and without sufficiently clear definitions in the relevant literature, which lead to confusion. That is why a terminological discussion to supplement the work presented here was deemed necessary: the most common terms are active, activated and aggressive versus passive, non-activated and non-aggressive sampling to discriminate between resuspending the air, or not, prior to sampling (Efthymiopoulos et al, 2021). Aktas et al. (2018a-c) Heinsohn (2007) and Swaebly and Christensen (1952) used the term *active* sampling to describe the collection of airborne particles after the disturbance of the air through artificial means, while *passive* sampling is used to describe the collection of particles from still air. Other researchers have used the same terms to differentiate between air sampling methods that require the creation of an artificial airflow for the collection of particles (e.g. impaction, liquid impingement, filtration and electrostatic precipitation) and methods that utilize gravitational settling (e.g. sedimentation), respectively (Grinshpun, 2010; Amato et al, 2018; Mainelis, 2019). To avoid common confusion over this overlap, in this paper we use the terms *activated (aggressive)* and *non-activated (non-aggressive)* to indicate still and perturbed indoor air conditions prior to sampling.

Terminological discourse aside, the boundary between what is considered to be still or actively-mixed air is also often unclear, which has particularly dire implications for studies focussed on exposure (Figure 2). Cahna et al., (2015) and Caballero et al., (2021) collected air samples for the assessment of the fungal contamination in classrooms while occupied by children. In another study by Shinohara et al., (2021), house dust samples were

collected from unoccupied houses in Japan, with no waiting between equipment set-up and sampling however. Aktas et al. (2018b) used a hand-held blower for 1 min per 10m<sup>2</sup> to resuspend particles prior to sampling. In all three cases, the environmental settings under which the air sampling was carried out were different and the findings not comparable. Although the use of a blower in the last example might be best placed to reach the “saturation point” and can be safely termed as an activated testing exercise (Figure 2), the other two cases sit within the wide spectrum of different levels of air stillness is still unclear.



**Figure 2: Illustration of non-activated (non-aggressive) and activated (aggressive) protocols**

## 1.2. Understanding the effect of activities and environmental context to the sampling readings

Many researchers have reported that the level and nature of the activities within a given indoor space do influence the extent and rate resuspension of bioaerosols, including the fungal matter (Table 2). The creation of artificial air currents due to the disturbance of the air’s steadiness was found to increase the concentration of the airborne fungi through resuspension, hence allowing the detection of particles that are otherwise not detectable (Rylander, 2015; Aktas et al., 2018b). However, how the readings obtained from non-activated and activated protocols differ is yet to be fully quantified. Importantly, the relevant literature is dominated by the use of non-activated protocols (e.g. Zorman and Jersek, 2008; Gent et al., 2012; Adams et al., 2013; Dallongeville et al., 2015). This is primarily on the claim that the collection of samples from still air can reduce reproducibility-related errors (e.g. Heinsohn, 2007), although a number of studies using both protocols comparatively suggest otherwise (e.g., Aktas et al., 2018a,b; Mukai et al., 2009).

There are a number of parameters which would define the nature and extent of activation and would impact on the readings in conjunction with it. The first and most immediate one is the changes in the **pattern and the speed of the inspector’s movement across space** which may lead to the resuspension of particles from different locations, potentially contaminated by fungi. Findings by Rylander (2015) indicate that the movement of inspectors inside residential buildings prior to testing increased the measured concentration of fungal biota by 2 times compared to when all movement was strictly prohibited. Small scale experiments and Computational Fluid Dynamics (CFD) tests carried out by Cao et al., (2017) indicated that human movement within a room could affect the dispersion of particles and that increase in the movement’s velocity is likely to lead to longer resuspension of fine particles (0.02-1µm).

**Table 2: Effect of the activation to the particle resuspension and fungal readings reported in the literature.**

<i>Research by</i>	<i>Context / Sampling and analysis method</i>	<i>Outcomes</i>
<i>Flannigan (1992)</i>	Field work / Impaction and colony enumeration	Suggested that during the occupied hours, the human activity led to a significant increase in the resuspension of fungal particles in the restaurant tested leading to an increase of the viable fungal counts ( <b>Approximately 16 times higher</b> than the readings during unoccupied hours)
<i>Rylander (2015)</i>	Field work/Filtration and NAHA measurements	Reported that that the indoor fungal levels (NAHA levels) in rooms in villas and apartments with no mechanical ventilation increased <b>up to 10 times</b> when an activated protocol using a blower was implemented compared to the readings when a non-activated protocol was selected.
<i>Aktas et al. (2018b)</i>	Field work/Impaction on Agar plates and colony enumeration & Filtration and NAHA detection	Reported an <b>up to 56 times</b> increase of the activated fungal readings compared to non-activated readings when testing was carried out in residential buildings in North London
<i>Mundt (2000)</i>	Laboratory experiments / Particle counters used to measure airborne particle concentrations	Reported an <b>up to 10 times</b> increase of the concentration of PM5 and PM10 when a person entered, and vigorously walked around the test room
<i>Mukai et al. (2009)</i>	Laboratory experiments / Particle counters used to measure airborne particle concentrations	Reported an <b>up to 10 times</b> increase of the relative resuspension rate of particles ranging from 0.5 to 20µm when the turbulence intensity and air velocity directly above three substrates (linoleum, metal, and carpet) increased by 23% and 20m/s respectively.
<i>Goldasteh et al. (2010)</i>	Laboratory experiments / Particle counters used to measure airborne particle concentrations	Reported 1) approximately <b>10 times</b> higher resuspension rates of 1-10µm particle from hardwood flooring and 2) approximately <b>2-3 times</b> higher resuspension rate of 1-10µm from linoleum flooring when airflow speed increased from 4.5 to 21 m/s in a laminar flow wind tunnel
<i>Napoli, Marcotrigiano &amp; Montagna (2012)</i>	Field work/Impaction and total viable counts (TVC enumeration)	Reported an increase of the mean TVC by approximately <b>7.5 times</b> during operations in hospital rooms in the Apulia Region in South-eastern Italy.
<i>Wang et al. (2021)</i>	Field work/ Particle counters used to measure airborne particle concentrations	Reported approximately <b>5.5 times</b> increase of average PM2.5 concentration when the occupancy density increased from 10 to 35 people in a classroom of Jinnan Campus of Nankai University

Another important parameter is the effect of the **room characteristics/condition**. Room characteristics, especially the level of cleanliness, can be connected to the amount of dust within a property and abiotic factors such as water and nutrient availability affecting the sporulation of certain fungal species (Buttner and Stetzenbach, 1993; Aktas et al, 2018a-c). In different properties, the quantity of dust and the fungi stored in it can vary. The volatility of fungi in dust can also change (Metz et al., 1979). Hence, the particle aerosolization rates may vary due to the different dust levels and volatility of fungal particles in every space and thus potential underestimation/overestimation issues regarding fungal readings may be risen. Another important concern regarding the detectability of fungal particles, especially the larger ones, is how quickly the airborne fungal levels return to the pre-activation levels. Activities may lead to different levels of resuspension of particles of different sizes. Allowing a long time to pass after the end of the resuspension may lead to the settlement of heavy particles before they are able to be collected through sampling (Tucker et al., 2007). Mundt (2000) suggests that large particles are expected to settle faster than small ones once indoor activities have stopped. To that end, the inability to specify the duration of time needed between the preparation of the indoor environment and the sampling, might lead to variations in the sampling results.

Finally, another concern with regards to both the comparability of the testing outcomes from the available testing procedures and their ability to assess the appropriate potential fungal reservoirs is the uncertain impact of the **hygrothermal conditions** on the readings. The relative humidity affects the fungal growth and sporulation (Arthurs and Thomas, 2001; Zhao and Shamoun, 2006; Money et al, 2016) directly, but can also indirectly influence the resuspension rates and the recovery efficiency of fungal particles indirectly. Depending on the

fungal species present the optimal relative humidity is likely to increase the spore production rate and conidial growth (Arthurs 2001; Zhao 2006; Money et al, 2016). However, the moisture content in the indoor environment and the indoor materials can also affect the adhesion of fungi on the indoor surfaces and may vary across different spaces (Osherov & May, 2001). In high humidity conditions, liquid molecules can be adsorbed on small-sized bioaerosols, including small-sized ( $>0.1\mu\text{m}$ ) fungal particles leading to an increase in the adhesion forces between the particles and the surfaces they come from in contact with (Baron and Kulkarni, 2011). As a result, the aerosolization rate of particles by implementing the testing methods may vary from case to case, thus influencing the sampling readings.

We therefore raise the following open research question: **What environmental settings prior to sampling ensure that the testing procedure is replicable, and lead to comparable results from different properties?** This paper aims to fill this gap by detailing the impact of the indoor environmental settings prior to the sampling on the testing outcomes through a critical review of the literature (Section 1), a series of laboratory experiments and a case study demonstration (Section 2) to then discuss research and knowledge gaps, and implications on the practice (Section 3) and draw conclusions (Section 4).

## 2.0 Methods

Experimental work and a case study testing demonstration were performed to examine the effect of the environmental settings to the resuspension rates of particles including fungal ones. Different protocols utilizing air activation via the resuspension of particles with a leaf blower were implemented in both segments of the study. While the experimental work aimed to test the effect of different activated protocols to the concentration of airborne particles with different sizes in a controlled environment, the case study was designed to examine the applicability and validity of the experimental outcomes in real life scenarios.

### 2.1. Experimental Work

In order to investigate, on a quantitative basis, the impact of the level of “activation” on the obtained readings, an experimental campaign was designed, and three sets of experiments were carried out. The experimental work aimed to identify the role of the blowing duration in the particle counts at different heights. The findings of this experimental work are then discussed in conjunction with the literature. For these experiments particle counter readings were chosen as a proxy for indoor mould testing outcomes. This was considered a robust approach since fungal particles can range in size from  $0.6\mu\text{m}$  –  $10\mu\text{m}$  (Claub, 2015) and associations between the airborne fungal concentration changes and particle counts have been previously reported in the literature (Agranovski et al., 2004; Brandl et al., 2008). No cultivation and aerosolization of fungal particles was carried out to minimize the risk of accidentally contaminating the test space.

The work was conducted in an environmental chamber ( $2.8\times 3\times 3\text{m}$ ) where particle counters were positioned at three different heights (Figure 3) - the readings from 6 particle size channels were analysed before and after activation. To ensure that dust or small particles cannot infiltrate the room through cracks and openings, the environmental chamber joints were entirely air-sealed with sealing tape typically used to perform blow-door tests. It is important to mention that the ventilation system in the chamber was not operated at any instance during these tests, and the ducts leading to the chamber were also sealed to avoid unintended disturbance in the air.

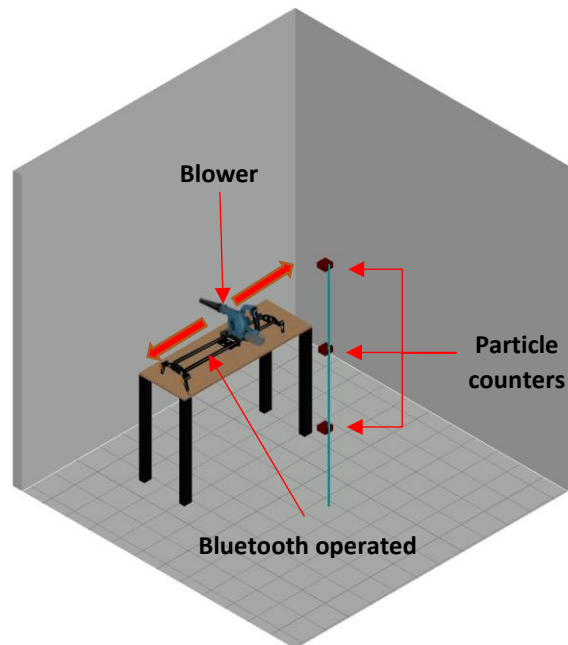
Three particle counters (PMS 5003 by Plantower, Sensitivity: 50% -  $0.3\mu\text{m}$ , 98% -  $0.5\mu\text{m}$  and larger, Resolution:  $1\mu\text{g}/\text{m}^3$ , work temperature: operating temperature  $-10\text{ }^{\circ}\text{C}$  to  $60\text{ }^{\circ}\text{C}$ , Humidity (work): 0-99%) were placed at heights of 0.75m, 1.5m and 2.25m on a vertical pole located in the centre of the room to monitor the variation of different sized particles with height. All three sensors were calibrated by the manufacturer prior to use.



**Figure 3: Experimental setup inside the environmental chamber.**

To implement activated protocols within the environmental chamber, a leaf blower (MODEL 100760 Merry Tools Air Leaf Dust Blower Electric Inflator) was used. The blower was placed on top of a Bluetooth operated slider (Neeer Motorized Camera Slider) and was connected to a wifi-enabled plug to ensure that it could be operated remotely, without entering the chamber. The blower was set at a fixed height (distance between nozzle end and wall =1.3m) and at a fixed distance from the wall (distance between nozzle end and wall =1.5m), and was able to move horizontally for a distance of 1m (Figure 4).

Experiments were conducted to examine the effect of 6 different blowing durations on the concentration of particles at the three different heights in the centre of the room. Tests were repeated three times (Series 1, 2, and 3 in Figure 6) to reduce the likelihood of anomalous results and increase the accuracy. The blowing durations were set to be 1min, 2min, 3min, 5min, 10min and 15min. During these periods, the blower was set to move horizontally from one end of the slider to the other, while blowing towards the wall at a rate of  $3.5 \text{ m}^3/\text{min}$ .



**Figure 4: Schematic representation of the experimental setup and the movement of the blower inside the environmental chamber.**

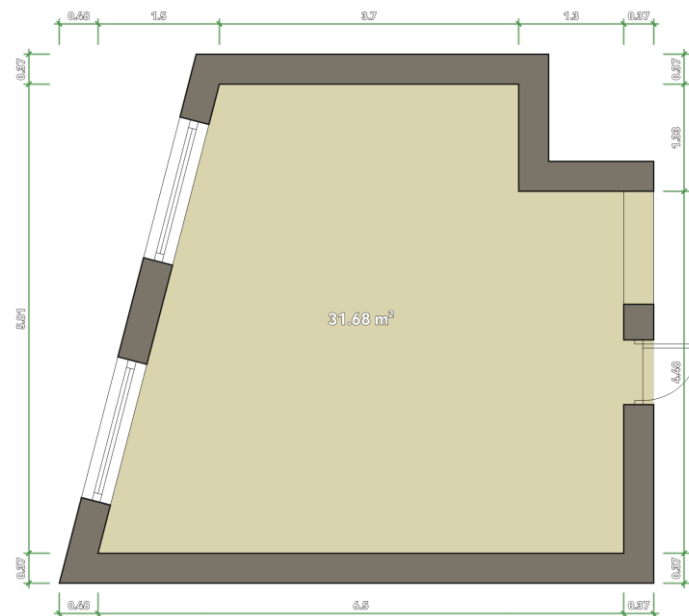
The experiments were conducted for a total of 20 days, and the chamber was closed and sealed until all experiments had been completed. Once the setup was complete, two days were allowed to pass before the experiments were initiated to ensure that all the resuspended dust due to the activities carried out for installation of the apparatus had settled down and different-sized particles' concentration did not significantly

change with time. The experiments were then initiated, and one experiment was carried out each day to ensure that the particles before the start of every experiment were in equilibrium. Within the context of these experiments, no dust was added inside the chamber, and the specific amount present inside the chamber was not known. However, knowing the exact amount of dust was not a prerequisite in this paper's context, as this study's aim was to identify how different activities would comparatively impact on the resuspension rates. Under no circumstance was anyone allowed to enter the room before the completion of all the experiments.

The particle counters were set up to start logging measurements 1 hour before the start of the activation and continued until 8 hours after it. The logging interval was 1 minute and the logs represent the average number of particles counted every second.

## 2.2. Case Study Testing

To validate the experimental data and understand the effect of different non-activated/activated protocols to air sampling fungal readings in real case scenarios, 6 experiments (Table 3) were conducted in the living room of an apartment in Eastern London (Figure 5). The property was located on the 3<sup>rd</sup> floor of a high-rise residential building and no visual signs of mould or dampness were detected anywhere inside. The occupants allowed the performance of the tests in the property and did not enter the apartment for the whole duration of the experiments to avoid affecting the sampling readings in any way.



**Figure 5: Floor plan of the case study living room**

The tests were designed to evaluate the effect of 4 different environmental settings to the sampling readings, which aimed to reflect different air-mixing (activation) states before sampling and involved carrying out what was deemed as no, light, medium and high-level activities (Table 3). The experiments for no, light and medium activity were performed once while the high activity case was repeated 3 times to investigate whether an activated protocol can be robust and reproducible.

The equipment was installed approximately 8.5 hours before the commencement of the first experiment. Upon completion of the setup, the living room doors and windows were closed and permission to enter the room was restricted to allow resuspended particles to settle and avoid accidentally aerosolizing any fungal particles from indoor surfaces. The first experiment was performed remotely using smart plugs to operate the pump and a DIY mechanism to uncover the filters' inlet. However, while for the first experiment all activities were restricted for more than 8 hours prior to sampling, the next 5 experiments were conducted consecutively with an approximately 10 minutes between each and the investigator entered the space and perform the sampling manually. For all experiments the investigator wore an FFP3 mask for respiratory protection purposes.



**Table 3: Description of activities carried out in every test before sampling**

<b>Experiment</b>	<b>Description</b>
<b>1</b>	<u>No activity</u> : The apartment remained unoccupied for more than 8 hours to allow any resuspended particles to settle before the commencement of the air sampling. Before initiating the sampling, the covers of the filters were removed by pulling a string from outside the living room
<b>2</b>	<u>Light activity</u> : The investigator walked inside the room for 2 minutes without moving objects, furniture or wiping dust off from any surface.
<b>3</b>	<u>Medium activity</u> : The investigator opened the window and walked inside the room for 5 minutes. Any other activity was restricted.
<b>4, 5 &amp; 6</b>	<u>High activity</u> : The investigator used a leaf blower (Makita BUB182) for approx. 1min per 10m <sup>2</sup> , to resuspend particles from any indoor surface (the distance between the investigator and the surface being blown was instructed to be 2m).

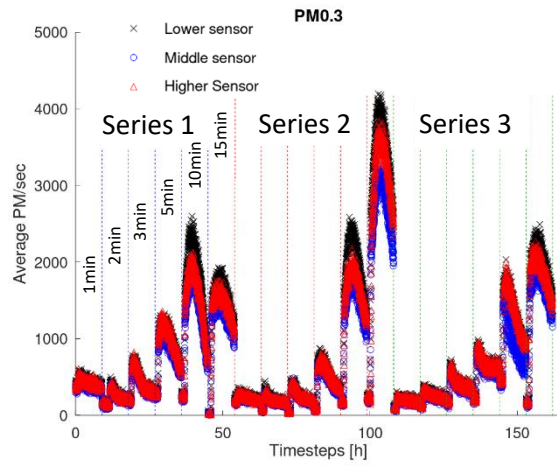
Air sampling via filtration was used to measure indoor fungal biomass and species after different levels of activities, representative of different activated/non-activated protocols. Two MCE-membrane filter (pore size 0.8µm) provided by Mycometer A/S, Denmark were used for each experiment, to quantify the fungal biomass present in the sampled air. One PTFE filter (pore size, 0.3µm) preloaded in a 37mm cassette was also used in every experiment to collect fungal-DNA and detect the presence of 17 targeted species in the sampled air. The sampling time and volumetric flow rate were selected as 10 min and 15 l/min respectively for both types of filters. Upon completion of the tests the MCE-membrane and PTFE filters were stored for 1 day in room temperature and a -80°C freezer in-house, and were sent and delivered to Mycometer A/S and Husetest ApS 1 and 2 days after the sampling, respectively.

The quantification of the fungal biomass was performed by Mycometer A/S using fluorogenic detection of different hydrolase activities ( $\beta$ -N-acetylhexosaminidase; EC 3.2.1.52; NAHA). Hydrolase substrate containing a fluorophore was contacted with the membrane MCE filters. The enzyme activity was determined by measuring the amount of fluorescence formed during the reaction (following the protocol of the manufacturer, Mycometer A/S). The fluorescence signals from the fungi and the total allergens were then extracted and reported in relative fluorescence units (RFU), along with the Fungal to Allergen Index (ratio of the fungi to allergen levels).

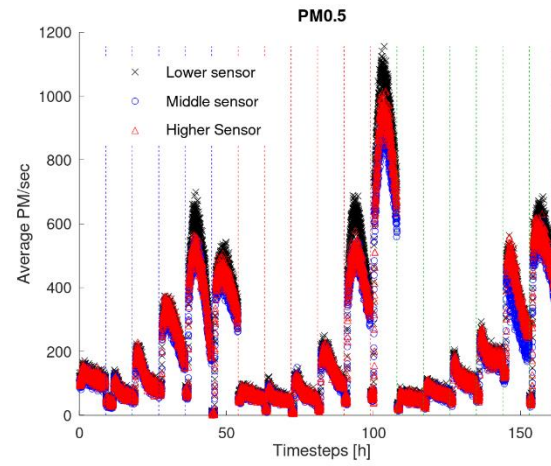
Species identification was performed by HouseTest ApS, Denmark through the extraction and amplification of DNA from the PTFE filters. Quantitative polymerase chain reaction (qPCR) analysis was performed using primers targeting the DNA of (a) 16 fungal species (*Acremonium strictum*, *Alternaria alternata*, *Aspergillus fumigatus*, *Aspergillus versicolor*, *Aspergillus niger*, *Chaetomium globosum*, *Cladosporium cladosporioides*, *Cladosporium herbarum*, *Cladosporium sphaerospermum*, *Penicillium chrysogenum*, *Penicillium expansum*, *Rhizopus stolonifer*, *Stachybotrys chartarum*, *Trichoderma viride*, *Ulocladium chartarum*, *Wallemia sebi*), and (b) three fungal groups (*Aspergillus glaucus* spp., *Mucor/Rhizopus* spp., *Penicillium/Aspergillus/Paecilomyces variotii* spp.). The internal transcribed spacer (ITS) region of the nuclear ribosomal repeat unit was targeted by the primer sequences used as described by Lu et al. (2021).

### **3.0 Results and Discussion**

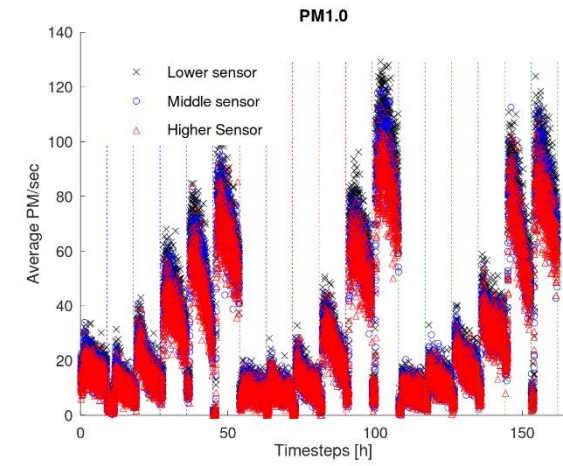
The outcomes of the experimental work are shown in Figure 6. Results indicate that the particle resuspension rates increase with the increase of the blowing time for all three sets of experiments, with up to X and Y times difference for small (PMa-b) and large particles (PMc-d), respectively, between sampling in still conditions and after 15 min of activation. Importantly, for blowing durations of more than 5min, the smaller particles (Particle size < PM1) did not return to the levels before the blowing was initiated, **even after 7 hours (Figure 6(a-c))**. However, the same cannot be stated for larger particles - for the same blowing duration, the pattern of the particle settlement becomes more unclear with the increase in the aerodynamic diameter (AD) (Figure 6(d-f)).



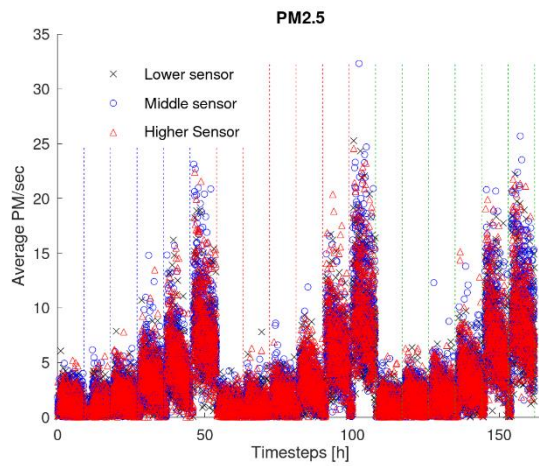
(a)



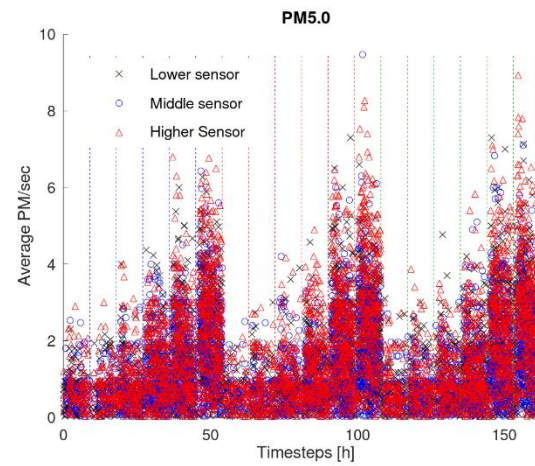
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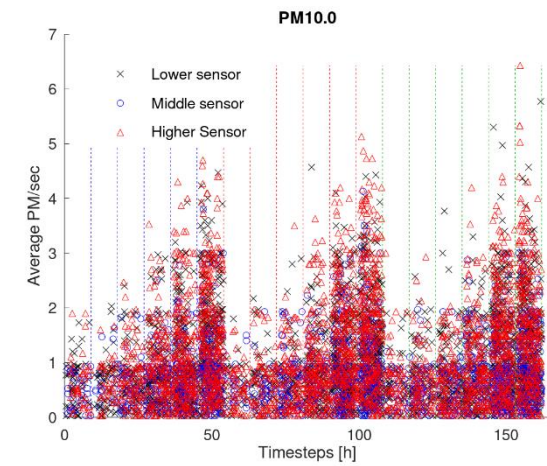
(c)



(d)



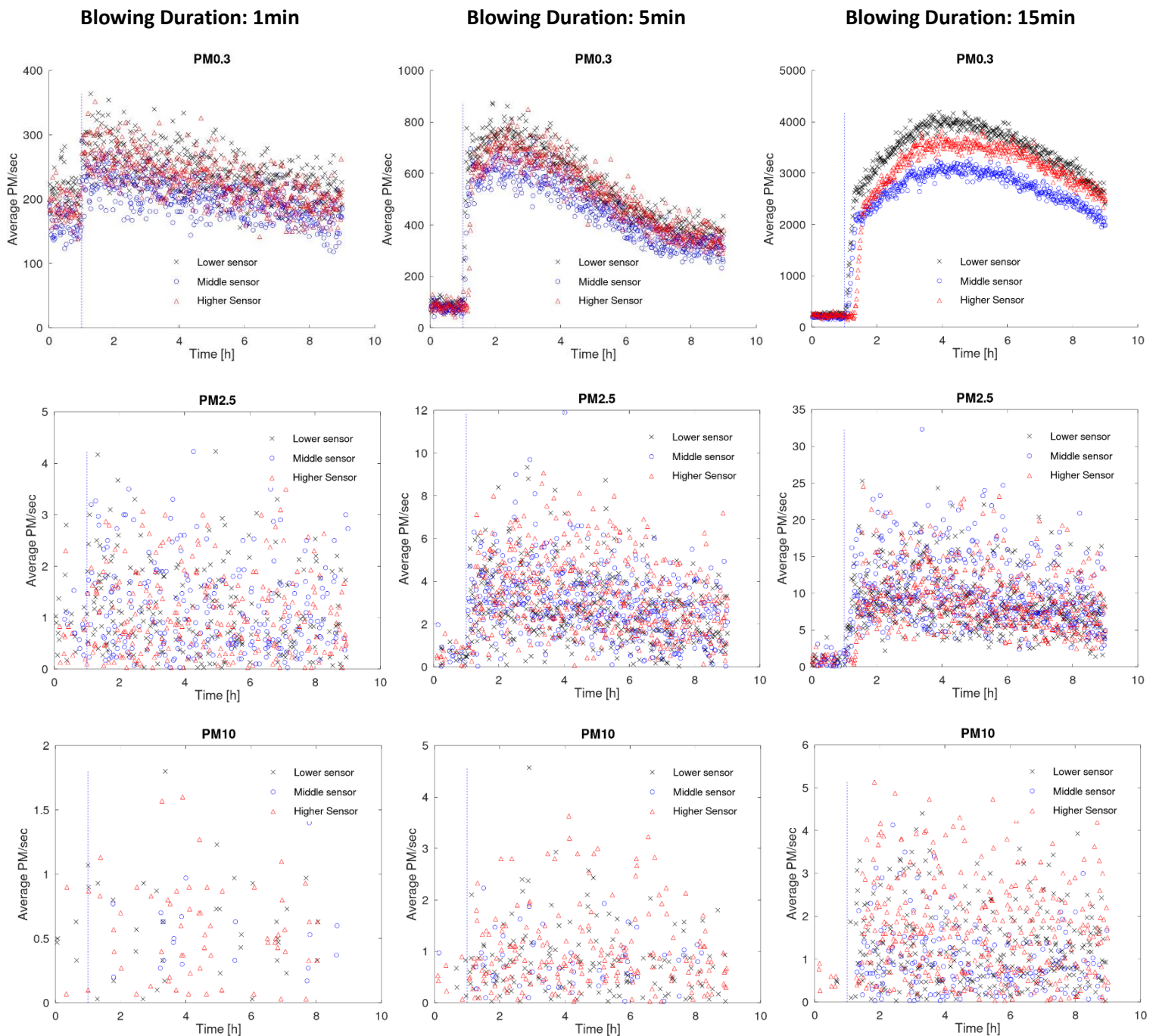
(e)



(f)

**Figure 6: Particle counts for (a) PM0.3, (b) PM0.5, (c) PM1.0, (d) PM2.5, (e) PM5.0, and (f) PM10. Different segments in each plot delineated in different colours indicate three repeated sets of each test. The findings are indicated for 1, 2, 3, 5, 10 and 15 min of blowing and for each of the three series as shown in (a).**

To better understand the pattern of the particle readings with the increase of the blowing duration, the average number of the PM0.3, PM2.5 and PM10 over 1 minute intervals were plotted against time for three cases of blowing duration: 1 min, 5 min and 15 min (Figure 7). As seen, higher blowing durations led to higher resuspension rates for all three size particles at any height.

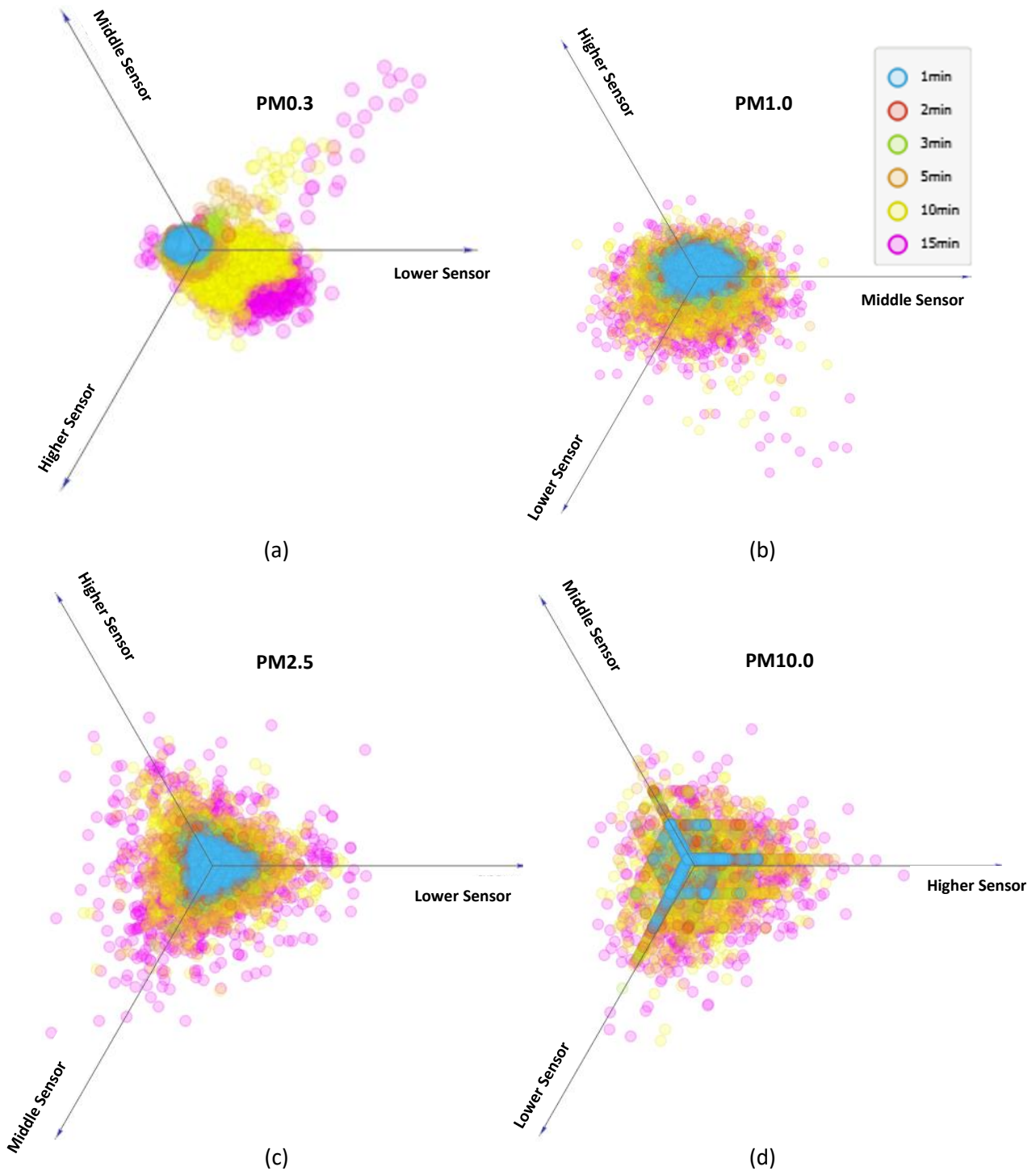


**Figure 7: Measurements of PM0.3, PM2.5 and PM10 at different heights for 3 blowing durations (1min, 5min, and 15min). The blue vertical dashed line represents the commencement of the blowing.**

The number of different sized particles measured at different heights was linearly projected to further study the effect of the blowing duration on the particle counts with height (Figure 8). The increase in the blowing duration affected the resuspension rate of both small and large particles. The difference in the particle numbers due to the change of the blowing duration is better captured by the lower particle sensor than the middle and higher one (Figure 8(a)). However, Figure 8(c-d) shows a more uniform increase in the levels of the particles when their AD is larger than  $1\mu\text{m}$  with increasing blowing durations compared to the levels of PM0.3 and PM0.5 (Figure 8

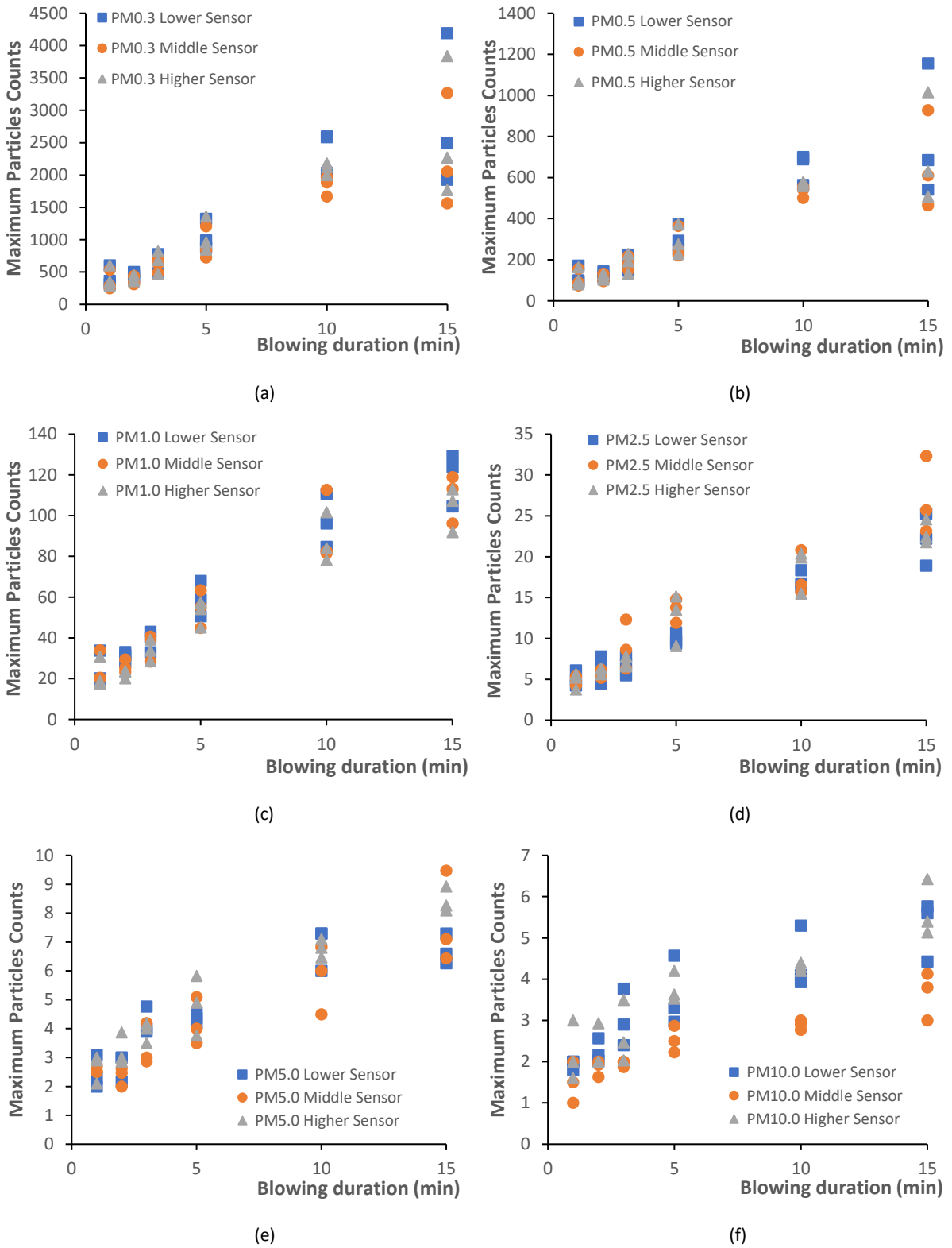


(a-b)). Higher levels of the small particles (PM0.3 and PM0.5) were measured by the lower sensor than the higher ones.



**Figure 8: Linear projection of same-sized particle readings measured at different heights**

The maximum number of particles measured at each height in every experiment were plotted against the blowing duration and are demonstrated in Figure 9(a-f). The coefficients of determination [ $R^2$ ] for all three series suggest that the maximum number of particles of all sizes is strongly correlated to the blowing duration with the PM1.0 and PM2.5. In conjunction with Figures 6 and 7, the increasing trend of the maximum values of particles with the prolongation of the blowing duration suggests that the increase of the activities duration prior to sampling leads to higher particle resuspension rates.



**Figure 9: Maximum levels of different sized particles with height for the three experimental series**

The Pearson's correlation coefficient was calculated to identify potential correlations between the number of different sized particles and the height, and a correlation matrix was created (Table 4). The results of the analysis indicate that the small particles (PM0.3 – PM2.5) correlate strongly with each other regardless of the height of the sensor ( $r > 0.6$ ). On the other hand, no strong correlations between the larger particles (PM5 and PM10) were identified with the highest value of  $r$  being 0.215 for PM5 particles measured by the higher and lower sensor and the lower  $R$ -value being the one for PM10 particles measured by the lower and higher sensor ( $r = 0.058$ ). It

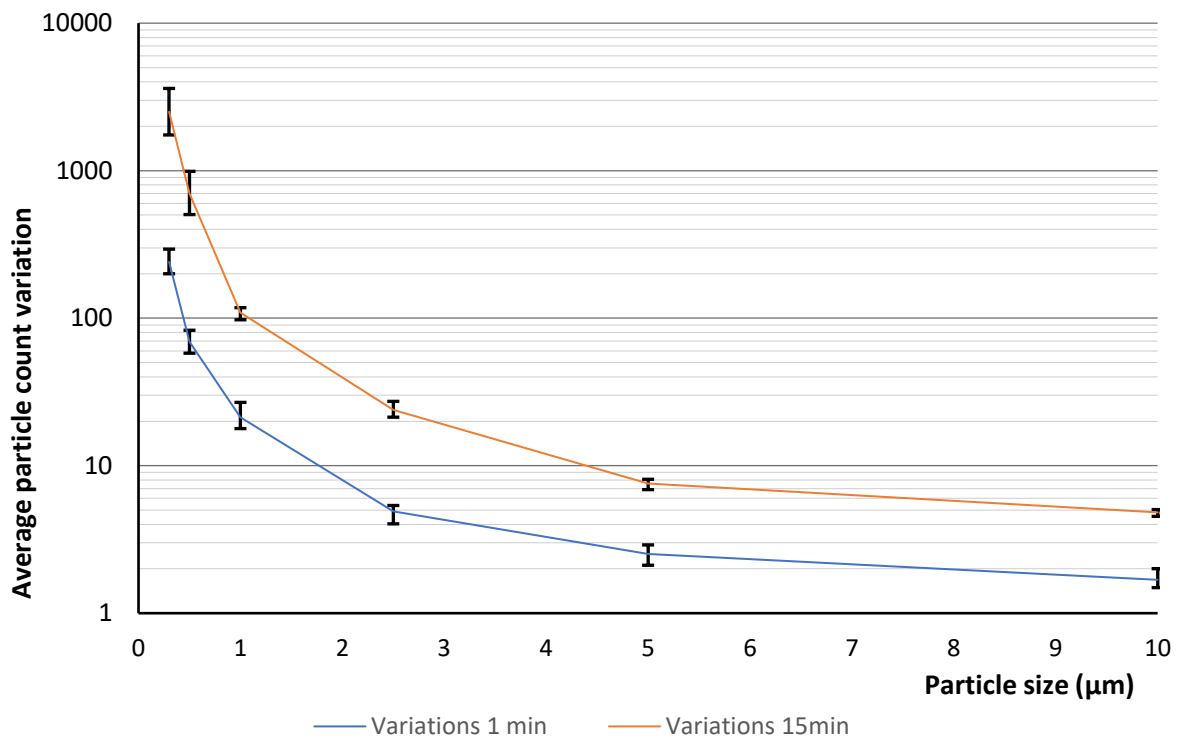
should be noted that in a previous study by Luoma and Batterman (2001), no significant variation was reported between readings of PM1 or smaller particles with height (particle counters placed at 0.4, 1.1, 1.8 m). On the other hand, the readings of particles ranging from PM5.0 – PM25 were shown to vary significantly with height. (Luoma and Batterman, 2001).

**Table 4: Correlation coefficient for same-sized particles measured at different heights**

	<i>PM0.3 Low Sensor</i>	<i>PM0.3 Mid Sensor</i>	<i>PM0.3 High Sensor</i>
<i>PM0.3 Low Sensor</i>	1.000		
<i>PM0.3 Mid Sensor</i>	0.993	1.000	
<i>PM0.3 High Sensor</i>	0.986	0.986	1.000
	<i>PM0.5 Low Sensor</i>	<i>PM0.5 Mid Sensor</i>	<i>PM0.5 High Sensor</i>
<i>PM0.5 Low Sensor</i>	1.000		
<i>PM0.5 Mid Sensor</i>	0.993	1.000	
<i>PM0.5 High Sensor</i>	0.985	0.986	1.000
	<i>PM1.0 Low Sensor</i>	<i>PM1.0 Mid Sensor</i>	<i>PM1.0 High Sensor</i>
<i>PM1.0 Low Sensor</i>	1.000		
<i>PM1.0 Mid Sensor</i>	0.964	1.000	
<i>PM1.0 High Sensor</i>	0.951	0.953	1.000
	<i>PM2.5 Low Sensor</i>	<i>PM2.5 Mid Sensor</i>	<i>PM2.5 High Sensor</i>
<i>PM2.5 Low Sensor</i>	1.000		
<i>PM2.5 Mid Sensor</i>	0.606	1.000	
<i>PM2.5 High Sensor</i>	0.606	0.611	1.000
	<i>PM5.0 Low Sensor</i>	<i>PM5.0 Mid Sensor</i>	<i>PM5.0 High Sensor</i>
<i>PM5.0 Low Sensor</i>	1.000		
<i>PM5.0 Mid Sensor</i>	0.142	1.000	
<i>PM5.0 High Sensor</i>	0.215	0.211	1.000
	<i>PM10 Low Sensor</i>	<i>PM10 Mid Sensor</i>	<i>PM10 High Sensor</i>
<i>PM10 Low Sensor</i>	1.000		
<i>PM10 Mid Sensor</i>	0.096	1.000	
<i>PM10 High Sensor</i>	0.058	0.090	1.000

The Levene test was used to examine the equality of the variance for the number of same sized particles measured by the same sensor during all three experiments. For a significance level  $p_{\text{value}}=0.05$  the tests have indicated no homogeneity of the variances between the three experimental series for all cases tested. This suggests that although the tests were repeated in the exact same way the variances of the readings are significantly different. These differences might be a result of the limited horizontal movement of the blower and the limited area being directly affected by the high air-velocity output. Nevertheless, this conforms with the literature outcomes in section 2 and adds extra value to the statement regarding the comparability concerns due to limited control over the activities and the testing conditions prior to sampling.

Further, to understand the extent of the activation's effect on the particle readings, the differences between the maximum and minimum particle counts captured by every sensor were averaged for all experiments carried out for 1min and 15min blowing durations and plotted against particle size in Figure 10. The blowing duration has led to an increase in the particle count variation for all particle sizes – with the difference being the largest for the smaller particles (> 4000 particles for PM0.3 and 15min blowing) and for PM10 particles (<7 particles) and blowing duration of 15min. This can indicate that the readings for the PM1.0 or smaller particles capture the blowing duration changes easier than the larger particle counts. Therefore, monitoring the small-sized particles (<PM1.0) could be a better proxy of the intensity of the activities carried out prior to sampling. It is important to mention that despite the prolonged blowing duration (15min), the variation in the large-particle readings (>PM2.5) was only slightly larger than the corresponding one when the blowing duration was 1min, indicating that the air activation might be more critical for the recovery efficiency of PM2.5 and larger particles during the sampling.



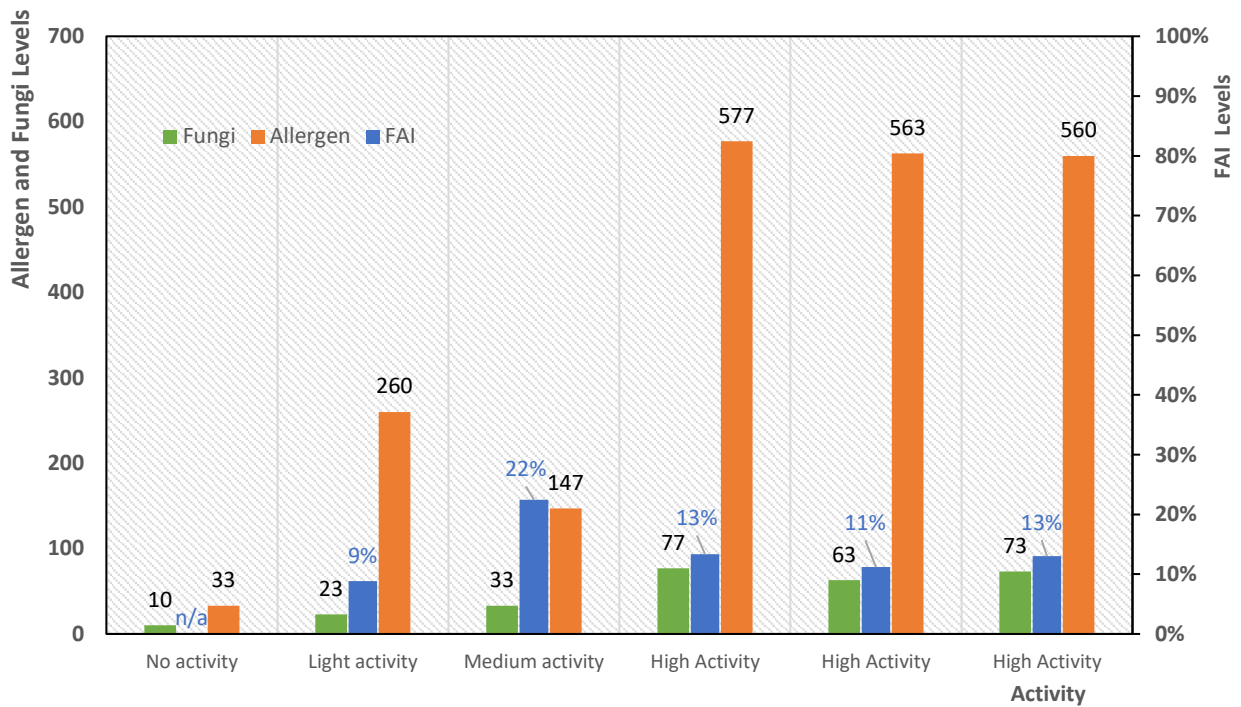
**Figure 10: Minimum and maximum readings' variations across the three series of testing for 1min and 15min blowing respectively**

### 3.1. Case study experimental outcomes

The Mycometer A/S method enabled the extraction of information with regards to three fungal growth indicators.

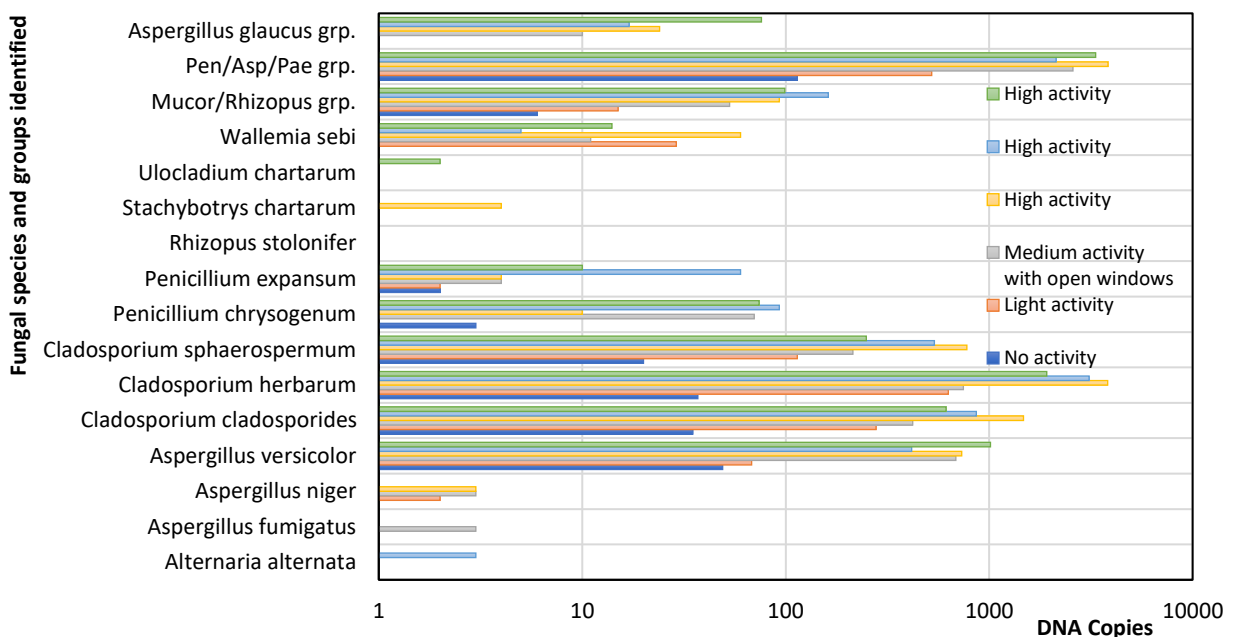
1. **The fungal levels:** Shows how many fungal particles have been detected in the 1st filter used during sampling.
2. **The allergen levels:** Shows the number of total allergens (Dust mites, pollen, fungi, pet dander, skin cells etc.) identified on the 2nd filter used during sampling.
3. **Fungal to Allergen Index (FAI):** measures the ratio between the two previous markers.

The fungal and allergen levels measured in the case study room indicated that the intensity of the activities prior to sampling has a noticeable impact to the testing readings (Figure 11). The first experiment (no activity) allowed the detection of very small concentrations of fungi (10RFU) and allergens (33RFU), while the results from the high activity experiments led to the capture of approximately 7 and 17 times more fungal and allergen particles in the MCE filters, respectively. It should be underlined that though the fungal levels increased when the activities intensified, the allergen levels did not follow the same trend. The allergen readings from the medium activity experiment were found to be lower than the corresponding ones from the light activity experiment. The underestimation of the allergen levels in the tested space might have been a result of the opening of the windows and the air exchange between a potentially poorer in allergens outdoor environment and an indoor environment with higher allergen levels. It is also worth noticing that the FAI indicators did not follow a particular trend with the increase of the activities' intensity. However, while the FAI ratios deviated noticeably in the first three experiments (no, light and medium activity), the deviation between the high activity ratios was rather minimal.



**Figure 11: Fungal to Allergen Index, fungal and allergen levels measured from all experiments**

Out of the 16 species that were targeted via the DNA amplification only 6 species were detected when activities prior to sampling were restricted completely (Figure 12). However, with the increase of the activity's intensity the number of identified species rose to 7, 10, 9, 9, 10 for the light, medium and high activity experiments respectively. Though the number of DNA clones for every group or species identified varied with the increase of the activity intensity, the fact that the samples from the no and light activity tests have captured 2-3 species less than the medium and high activity tests might mean that non-activated protocols might be underestimating fungal activity in the rooms. In addition, the fact that *Aspergillus fumigatus* was only detected in very small concentration in the sample from the medium activity test where the windows were open, raises concerns as to whether the species richness (number of identified species; s) in the sample was affected by the air exchange between the indoor and outdoor environment.



**Figure 12: Fungal species identified from every case study test.**



The fungal species richness (s), evenness (E) and diversity (H) were calculated to assess biodiversity in the samples collected. The Smith and Wilson evenness index (Equation 1) (Smith & Wilson, 1996) was used to determine how equally is the species abundance distributed in the samples while the Shannon & Wiener diversity index (SDI; Equation 2) (Shannon & Weaver, 1949) was used as a metric to study the diversity of species in the case study samples. These indicators have been widely used in studies focusing in biological diversity and ecological monitoring, and have been found to produce invaluable information regarding the distribution of species with different occurrences in samples. The Smith and Wilson evenness index was selected and used instead of any other evenness metric, due to its' low dependence to the species richness (Smith & Wilson, 1996; Goudarzian & Erfanfard, 2017) which could have led to misleading results, given the low number of targeted species, if other indicators were to be used. The biodiversity indicators have been summarized in Table 5.

$$E = 1 - \left[ \frac{2}{\pi \arctan \left\{ \frac{\sum_{i=1}^S (\ln n_i - \sum_{j=1}^S \ln n_j / S)^2 / S}{S} \right\}} \right] \quad [\text{eq.1}]$$

$$H = -\sum [P_i \ln(P_i)] \quad [\text{eq.2}]$$

**Table 5: Species richness, diversity and evenness indicators for the case study samples.**

<b>Experiment</b>	<b>Species Richness (s)</b>	<b>SDI (H)</b>	<b>Evenness (E)</b>
<i>No activity</i>	6	1.468	0.354
<i>Light activity</i>	9	1.201	0.109
<i>Medium activity</i>	10	1.451	0.107
<i>High activity</i>	9	1.204	0.085
<i>High activity</i>	9	1.184	0.090
<i>High activity</i>	10	1.285	0.083

The biodiversity indicators show that the activities carried out prior to sampling can have noticeable effects on an investigator's understanding of the species distribution in the tested spaces. The evenness index was found to have a decreasing trend with the increase of the activities' intensity which may indicate an increase in the difference in species' abundance due to higher fungal particle resuspension rates when more intensive activities are carried out. However, no particular trend was detected for the SDI with the increase in the activities' intensity. The unclear pattern for the SDI can be attributed to the fact that the SDI is positively affected by the increase of both the species richness and evenness of the species' abundance. Therefore, the increase in the number of the identified species and the decrease in their abundance evenness with the intensification of the activities had an unclear effect on the SDI results.

While the SDI and evenness values measured from the high activity samples vary by up to 0.11 and 0.07, respectively, the corresponding values for the no, light and medium activity experiments varied by up to 0.267 and 0.247 respectively. The high activity biodiversity indicators showcase that the species diversity and abundance in the samples had noticeably smaller fluctuations than any other given set from the case study experiments. Therefore, failing to well-define the environmental settings under which the sampling is conducted may have serious implications on an inspector's understanding of the species distribution in the tested spaces.

### **3.2. Discussion**

The experimental work has shown that the particle readings, regardless of the particle size, follow an increasing trend with the increase of the blowing duration. However, the impact of air activation on larger particle readings (>PM2.5) was considerably weaker than the effect of the smaller ones (<PM2.5). Previous studies (Quian and Ferro, 2008; Wang et al., 2021) have shown that the indoor air velocity close to a dust reservoir and the

activation method used can affect the particle resuspension rates. Still, they have also stressed that large particles might be more inert to activation due to their size and the strong adhesion bonds between them and the surfaces they are attached to. Therefore, investigators should consider that a strict-non-activated protocol might seriously underestimate the particle intensity with important implications in relation to the indoor fungal levels and the health impact.

The particle counters' low sensitivity for PM<sub>0.3</sub> readings (50% sensitivity) indicates that the readings of this channel might not be an accurate proxy for the evaluation of the indoor activities before sampling. Still, they might be able to show a trend for the PM<sub>0.3</sub> concentration. Given also that the majority of fungal particles will range between 1.0 µm AD and 3.2 µm AD (Claub, 2015), it can be stated that the PM<sub>0.3</sub> readings would not be a suitable metric for the indoor fungal resuspension rates. On the other hand, the high sensitivity (>98%) of the sensor for particles larger than PM<sub>0.5</sub> and the considerably higher particle count differences for smaller-sized particles (<PM<sub>1.0</sub>) with the increase of the blowing duration (Figure 10) indicates that the PM<sub>0.5</sub> and PM<sub>1.0</sub> readings reported here could be the most appropriate indicators of the activities carried out before sampling.

The high correlation coefficients between readings obtained at different heights for <PM<sub>2.5</sub> (Table 4) indicate that the height at which the sensors are placed will not significantly affect their ability to capture the concentration changes of particles. This conforms with previous studies showing that regardless of the sensor's height, unlike the coarser particles (>PM<sub>5.0</sub>), the smaller airborne particle counts will follow similar trends due to their ability to spread throughout an indoor space unrestricted of their size (Montoya & Hildemann, 2005). However, the same cannot be stated for larger particles (>PM<sub>2.5</sub>) where the low correlation coefficients suggest that the height of the sensor had led to noticeable differences in the particle count changes with time.

When looking upon the case study outcomes, the small deviations between the FAI, fungi and allergen levels from the high activity tests, especially when compared to the deviations between the other cases, suggest that activated protocols, if well-defined, can be reproducible and provide robust fungal measurements. The results also indicate that the increase of the activity intensity prior to sampling affects the fungal readings and raise concerns with regards to how activities like the opening of windows might lead to measurements that may not be representative of the indoor fungal activity due to the mixing of indoor and outdoor air before the sampling. The increase of the fungal readings with the escalation of the activities, comes in agreement with previous studies by Rylander (2015) and Aktas et. al. (2018b) who showed that the use of a blower prior to sampling led to an up to 10 and 56 times increase in the testing outcomes compared to the no and low activity readings respectively.

It is worth noting that the intensification of activities prior to sampling has allowed the detection of more species than the ones captured from the no activity case. However, while the low activity experiment allowed the identification of an additional species in the sample all other experiments led to the detection of 3 or more species. This is in broad agreement with the hypothesis that air activation can allow the aerosolization and detection of mould particles that develop stronger bonding forces with the substrate materials and would not be identified via non-activated protocols were carried out prior to sampling.

## **4.0 Conclusions**

Researchers should give attention to the conditions under which the indoor fungal testing is carried out. Outcomes of non-activated and activated testing do differ, therefore indoor fungal levels cannot be evaluated or benchmarked unless uniformity is brought to the pre-sampling conditions through a well-established testing protocol. In the case of activated sampling, disturbing the air's stillness may increase the concentration of airborne fungal fragments and spores, and thus could lead to the detection of fungi that could otherwise be undetectable. The experimental work carried out in the context of this study suggests that the use of a blower and the increase of the blowing duration could lead to higher particle resuspension which in real case scenarios could lead to higher detectability of particles ranging from 0.3 to 10 µm, as supported also by the case study testing we present here. It should be underlined that the findings suggest that the smaller particle (PM<sub>0.3</sub> to PM<sub>1.0</sub>) readings may be able to capture the different levels of activity better than the readings from larger particles (PM<sub>2.5</sub> and PM<sub>5.0</sub>).

Therefore, the selection of an activated protocol is of critical importance for the large particles' collection efficiency. Furthermore, the experimental study presented here suggests that the sampling height is another testing protocol variable that the larger particles are more sensitive to, which should bring some uniformity for comparability purposes. At the same time, the case study testing has shown that the intensification of the activities carried out prior to sampling allows the identification of more species than non-activated protocols. However, activities that could lead to an increase/decrease of contaminants indoors, such as the opening of windows, can have unclear effects on the fungal measurements and thus produce misleading results with regard to the assessment of indoor fungal activity.

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#### **5.0 References**

- Adams, R.I. et al. (2013) 'Dispersal in microbes: fungi in indoor air are dominated by outdoor air and show dispersal limitation at short distances', *The ISME Journal*, 7(7), pp. 1262–1273. doi:10.1038/ismej.2013.28.
- Agranovski, V. et al. (2004) 'Size-selective assessment of airborne particles in swine confinement building with the UVAPS', *Atmospheric Environment*, 38(23), pp. 3893–3901. doi:10.1016/j.atmosenv.2004.02.058.
- Aktas, Y. D., et al. (2018a). Indoor Mould Testing and Benchmarking: Public Report. (February), 1–16.
- Aktas, Y. D. et al. (2018b) 'Surface and passive/active air mould sampling: A testing exercise in a North London housing estate', *Science of the Total Environment*, 43, pp. 1631–1643.
- Aktaş, Y.D., Shi, J., Blades, N., D'Ayala, D. (2018c) Indoor mould testing in a historic building: Blickling Hall, *Heritage Science* 6(51), pp. 1-9, doi: 10.1186/s40494-018-0218-x
- Aktas, Y.D. et al. (2020) 'Normal background levels of air and surface mould reserve in English residential building stock: a preliminary study towards benchmarks based on NAHA measurements', *UCL Open Environment*, 2(1). doi:10.14324/111.444/ucloe.000005.
- Amato, P., et al. (2018). Sampling Techniques. In A.-M. Delort & P. Amato (Eds.), *Microbiology of Aerosols* (1st ed.). Hoboken: John Wiley & Sons.
- Andersson, M.A. et al. (1997) 'Bacteria, molds, and toxins in water-damaged building materials', *Applied and Environmental Microbiology*, 63(2), pp. 387–393. doi:10.1128/aem.63.2.387-393.1997.
- Andersen, B., et al. (2021). 'A Pilot Study on Baseline Fungi and Moisture Indicator Fungi in Danish Homes'. *Journal of Fungi* 7, no. 2: 71. <https://doi.org/10.3390/jof7020071>.
- Arthurs, S. and Thomas, M.B. (2001) 'Effects of Temperature and Relative Humidity on Sporulation of *Metarhizium anisopliae* var. *acridum* in Mycosed Cadavers of *Schistocerca gregaria*', *Journal of Invertebrate Pathology*, 78(2), pp. 59–65. doi:10.1006/jipa.2001.5050.
- Baron, P.A., Kulkarni, P. and Willeke, K. (2011) 'Aerosol measurement: principles, techniques, and applications.' 3rd ed. Hoboken, N.J: Wiley.
- Brandl, H., et al. (2008). 'Short-term dynamic patterns of bioaerosol generation and displacement in an indoor environment.' *Aerobiologia*, 24(4), 203-209.
- BSI, (2014). 'BS ISO 16000-19: 2014 Indoor Air: Sampling strategy for moulds', Brussels: CEN.
- Buttner, M.P. and Stetzenbach, L.D. (1993) 'Monitoring Airborne Fungal Spores in an Experimental Indoor Environment To Evaluate Sampling Methods and the Effects of Human Activity on Air Sampling', *APPL. ENVIRON. MICROBIOL.*, 59, p. 8.

- Canha, N. et al. (2015) 'Assessment of bioaerosols in urban and rural primary schools using passive and active sampling methodologies', *Archives of Environmental Protection*, 41(4), pp. 11–22. doi:10.1515/aep-2015-0034.
- Cao, S.-J. et al. (2017) 'Study on the impacts of human walking on indoor particles dispersion using momentum theory method', *Building and Environment*, 126, pp. 195–206. doi:10.1016/j.buildenv.2017.10.001.
- Clauß, M. (2015) 'Particle size distribution of airborne microorganisms in the environment - a review', *Landbauforschung - applied agricultural and forestry research*, (2/2015), pp. 77–100. doi:10.3220/LBF1444216736000.
- Dallongeville, A. et al. (2015) 'Concentration and determinants of molds and allergens in indoor air and house dust of French dwellings', *Science of The Total Environment*, 536, pp. 964–972. doi:10.1016/j.scitotenv.2015.06.039.
- Efthymiopoulos, S., Aktas, Y., Altamirano, H. (2021) 'Air sampling and analysis of indoor fungi: a critical review of passive (non-activated) and active (activated) sampling'. 1st International Conference on Moisture in Buildings 2021 (ICMB21). DOI: 10.14293/ICMB210021
- Flannigan, B. (1992) 'Indoor microbiological pollutants-sources, species, characterisation and evaluation.' In *Chemical, Microbiological, Health and Comfort Aspects of Indoor Air Quality* Gate of the Art in SBS (Edited by Knoppel, H. and Wolkoff, P.), pp. 73398. Kluwer, Dordrecht.
- Flannigan, B. (1997) 'Air sampling for fungi in indoor environments', *Journal of Aerosol Science*, 28(3), pp. 381–392. doi:10.1016/S0021-8502(96)00441-7.
- Gent, J.F. et al. (2012) 'Household mold and dust allergens: Exposure, sensitization and childhood asthma morbidity', *Environmental Research*, 118, pp. 86–93. doi:10.1016/j.envres.2012.07.005.
- Goldasteh, I., Ahmadi, G. and Ferro, A. (2010) 'Effect of Air Flow on Dust Particles Resuspension From Common Flooring', in *ASME 2010 3rd Joint US-European Fluids Engineering Summer Meeting: Volume 1, Symposia – Parts A, B, and C. ASME 2010 3rd Joint US-European Fluids Engineering Summer Meeting collocated with 8th International Conference on Nanochannels, Microchannels, and Minichannels*, Montreal, Quebec, Canada: ASMEDC, pp. 2797–2800. doi:10.1115/FEDSM-ICNMM2010-30596.
- Górny RL (2004) Filamentous microorganisms and their fragments in indoor air – a review. *Ann Agric Environ Med*, 11, pp. 185–197.
- Gots, R.E., Layton, N.J. and Pirages, S.W. (2003) 'Indoor Health: Background Levels of Fungi', *AIHA Journal*, 64(4), pp. 427–438. doi:10.1080/15428110308984836.
- Grinshpun, S. A. (2010) *Biological Aerosols*. In *Aerosols - Science and Technology*. Edited by I. Agranovski. Weinheim: Wiley-VCH.
- Gutarowska, B., Skóra, J. and Pielech-Przybylska, K. (2015) 'Evaluation of ergosterol content in the air of various environments', *Aerobiologia*, 31(1), pp. 33–44. doi:10.1007/s10453-014-9344-4.
- Heinsohn, P. A. (2007). *Conducting building mold investigations*. In C. S. Yang & P. A. Heinsohn (Eds.), *Sampling and Analysis of Indoor Microorganisms*. Pacifica, Calif: Wiley Interscience
- Luoma, M. and Batterman, S.A. (2001) 'Characterization of Particulate Emissions from Occupant Activities in Offices: Characterization of Particulate Emissions from Occupant Activities in Offices', *Indoor Air*, 11(1), pp. 35–48. doi:10.1034/j.1600-0668.2001.011001035.x.
- Lu, R., et al. (2021) 'Species of Fungi and Pollen in the PM1 and the Inhalable Fraction of Indoor Air in Homes.' *Atmosphere*. 12 (3), 404. doi:10.3390/atmos12030404
- Mainelis, G. (2019) 'Bioaerosol sampling: Classical approaches, advances, and perspectives', *Aerosol Science and Technology*. Taylor & Francis, 0(0), pp. 1–24.
- Metz, B., Kossen, N.W.F. and Suijdam, J.C. (1979) 'The rheology of mould suspensions', in *Advances in Biochemical Engineering*, Volume 11. Berlin, Heidelberg: Springer Berlin Heidelberg (Advances in Biochemical Engineering/Biotechnology), pp. 103–156. doi:10.1007/3-540-08990-X\_24.
- 'Microorganisms in home and indoor work environments: diversity, health impacts, investigation and control' (2002) *Choice Reviews Online*, 40(04), pp. 40-2201-40-2201. doi:10.5860/CHOICE.40-2201.

- Money, N.P. (2016) 'Spore Production, Discharge, and Dispersal', in *The Fungi*. Elsevier, pp. 67–97. doi:10.1016/B978-0-12-382034-1.00003-7.
- Montoya, L.D. and Hildemann, L.M. (2005) 'Size distributions and height variations of airborne particulate matter and cat allergen indoors immediately following dust-disturbing activities', *Journal of Aerosol Science*, 36(5–6), pp. 735–749. doi:10.1016/j.jaerosci.2004.11.004.
- Mukai, C., Siegel, J.A. and Novoselac, A. (2009) 'Impact of Airflow Characteristics on Particle Resuspension from Indoor Surfaces', *Aerosol Science and Technology*, 43(10), pp. 1022–1032. doi:10.1080/02786820903131073.
- Mundt, E., (2022). 'PARTICLES AND DISPLACEMENT VENTILATION.' In: H. Awbi, ed., *Air Distribution in Rooms, (ROOMVENT 2000)*, 1st ed. Reading: Elsevier Science Ltd, pp.737-742.
- Napoli, C., Marcotrigiano, V. and Montagna, M.T. (2012) 'Air sampling procedures to evaluate microbial contamination: a comparison between active and passive methods in operating theatres', *BMC Public Health*, 12(1), p. 594. doi:10.1186/1471-2458-12-594.
- Nastasi, N. et al. (2020) 'Morphology and quantification of fungal growth in residential dust and carpets', *Building and Environment*, 174, p. 106774. doi:10.1016/j.buildenv.2020.106774.
- Nieto-Caballero, M. et al. (2022) 'Aerosol fluorescence, airborne hexosaminidase, and quantitative genomics distinguish reductions in airborne fungal loads following major school renovations', *Indoor Air*, 32(1). doi:10.1111/ina.12975.
- Osharov, N. and May, G.S. (2001) 'The molecular mechanisms of conidial germination', *FEMS Microbiology Letters*, 199(2), pp. 153–160. doi:10.1111/j.1574-6968.2001.tb10667.x.
- Qian, J. and Ferro, A.R. (2008) 'Resuspension of Dust Particles in a Chamber and Associated Environmental Factors', *Aerosol Science and Technology*, 42(7), pp. 566–578. doi:10.1080/02786820802220274.
- Rao, C.Y., Burge, H.A. and Chang, J.C.S. (1996) 'Review of Quantitative Standards and Guidelines for Fungi in Indoor Air', *Journal of the Air & Waste Management Association*, 46(9), pp. 899–908. doi:10.1080/10473289.1996.10467526.
- Rylander, R. (2015) 'β-N-Acetylhexosaminidase (NAHA) as a Marker of Fungal Cell Biomass – Storage Stability and Relation to β-Glucan', *International Journal of Environmental Monitoring and Analysis*, 3(4), p. 205.
- Shannon, C.E. & Weaver, W. (1949) *The Mathematical Theory of Communication*. University of Illinois Press, Urbana.
- Shinohara, N. (2021) 'Comparison of DNA sequencing and morphological identification techniques to characterize environmental fungal communities', *Scientific Reports*, p. 8.
- Singh, J. (1999) 'Review: Dry Rot and Other Wood-Destroying Fungi: Their Occurrence, Biology, Pathology and Control', *Indoor and Built Environment*, 8(1), pp. 3–20. doi:10.1177/1420326X9900800102.
- Smith, B. and Wilson, J. B. (1996). 'A consumer's guide to evenness indices.' - *Oikos* 76: 70-82.
- Swaebly, M. A. and Christensen, C. M. (1952) 'Molds in house dust, furniture stuffing, and in the air within homes', *Journal of Allergy*, 23(4), pp. 370–374.
- Terčelj, M., et al. (2011). 'Fungal exposure in homes of patients with sarcoidosis - An environmental exposure study.' *Environmental Health: A Global Access Science Source*, 10(1). <https://doi.org/10.1186/1476-069X-10-8>
- Tucker, K. et al. (2007) 'Biomechanics of conidial dispersal in the toxic mold *Stachybotrys chartarum*', *Fungal Genetics and Biology*, 44(7), pp. 641–647. doi:10.1016/j.fgb.2006.12.007.
- Wang, B. et al. (2021) 'Experiments and simulations of human walking-induced particulate matter resuspension in indoor environments', *Journal of Cleaner Production*, 295, p. 126488. doi:10.1016/j.jclepro.2021.126488.
- Yamamoto, N. et al. (2015) 'Indoor Emissions as a Primary Source of Airborne Allergenic Fungal Particles in Classrooms', *Environmental Science & Technology*, 49(8), pp. 5098–5106. doi:10.1021/es506165z.

- Zhao, S. and Shamoun, S.F. (2006) 'The effects of culture media, solid substrates, and relative humidity on growth, sporulation and conidial discharge of *Valdensinia heterodoxa*', *Mycological Research*, 110(11), pp. 1340–1346. doi:10.1016/j.mycres.2006.08.001.
- Zorman, T. and Jeršek, B. (2008) 'Assessment of Bioaerosol Concentrations in Different Indoor Environments', *Indoor and Built Environment*, 17(2), pp. 155–163. doi:10.1177/1420326X08089251.
- World Health Organization Europe (2009) 'Dampness and mould', WHO guidelines for indoor air quality: dampness and mould, pp. 1–228.
- World Health Organization Europe (2013) 'Combined or multiple exposure to health stressors in indoor built environments', (Edited by Dimosthenis A. Sarigiannis), Bonn, Germany